



INTERNATIONAL JOURNAL OF PHARMACY & LIFE SCIENCES

A review on marine immunomodulators

Vikrant Arya* and Vivek Kumar Gupta

Department of Pharmacognosy,
Amar Shaheed Baba Ajit Singh Jujhar Singh Memorial College of Pharmacy,
Bela, Ropar, (Punjab) - India.

Abstract

Many natural products from marine sources are endowed with promising immunomodulating activities, thus representing invaluable leads in the drug discovery. The aim of this review is to highlight the work on natural immunomodulators from marine origin with around 35 naturally available marine sources have been explained. This review also discusses various chemical constituents responsible for immunomodulatory action along with their mechanism involved in immunomodulation. This work stimulates the researchers for further work on the marine immunomodulators.

Key-Words: Marine, Immunomodulatory, Immune system.

Introduction

Marine natural products

Natural products as crude materials with efficacy against various diseases have been selected by humans over many generations of practical experience. Such experiential evaluation is different from the scientific evaluation of western medicines and is underestimated sometimes. However many effective medicines, including as morphine, aspirin, atropine, ephedrine, reserpine and digitoxin were developed from natural products¹.

Natural products have always played an important role in medicine and in particular, marine metabolites have increasingly become major players in recent drug discovery. It depends on the structurally unique molecules produced by marine organisms, containing a significant number of stereogenic centres and halogen atoms, the latter introduced by metabolic processes of the inorganic species present in sea waters².

Recent studies of marine organisms have focused on their potential applications, particularly in the treatment of human diseases. Several marine natural products are currently in pre-clinical and clinical evaluation, others show promising biological activities³.

New trends in drug discovery from natural sources emphasize on investigation of the marine ecosystem to explore numerous complex and novel chemical entities. These entities are the sources of new leads for treatment of many diseases such as cancer, AIDS, inflammatory conditions and a large variety of viral, bacterial and fungal diseases⁴.

Marine chemicals have novel structures with pronounced biological activity and pharmacology. The study of such chemicals therefore is promising⁵. The potential for marine natural products as pharmaceutical was first developed in the 1950s which led to two marine derived pharmaceuticals that are still in use today. About 15,000 natural products have been described and about 30% of these natural products have been isolated from sponges Ara-C is an anti-cancer drug (used against non-Hodgkin's lymphoma and acute myelocytic leukemia) and Ara-A used as an antiviral drug for treating herpes⁶.

Immunomodulators

These are biological or synthetic substances, which can stimulate, suppress or modulate any of the immune system including both adaptive and innate arms of the immune response. Clinically immunomodulators can be classified into following three categories.

Immunoadjuvants: These agents are used for enhancing vaccines efficacy and therefore, could be considered specific immune stimulants⁷, example in this regard is of Freund's adjuvant⁸. The immunoadjuvants hold the promise of being the true modulators of immune response. It has proposed to

* Corresponding Author

E mail: arya.vikrant30@gmail.com

Mob. 09736105832

exploit them for selecting between cellular and humoral, Th1 (helper T1 cells) and Th2, (helper T2 cells) immunoprotective and immunodestructive, and reagenic (IgE) versus immunoglobulin G (IgG) type of immune responses, which poses to be a real challenge to vaccine designers⁷.

Immunostimulants: These agents are inherently non-specific in nature as they envisaged to enhance body's resistance against infection. They can act through innate immune response and through adaptive immune response. In healthy individuals the immunostimulants are expected to serve as prophylactic and promoter agents i.e. as immunopotentiators by enhancing basic level of immune response, and in the individual with impairment of immune response as immunotherapeutic agents⁹.

Immunosuppressants: These are a structurally and functionally heterogeneous group of drugs, which are often concomitantly administered in combination regimens to treat various types of organ transplant rejection and autoimmune diseases¹⁰. Various marine sources used as immunomodulatory agents have been mentioned in table 1 given below.

Chemistry of marine immunomodulators¹¹⁻³⁷

Wide ranges of phytoconstituents as shown in figure 1. were responsible for immunomodulatory activity includes Cyclic depsipeptide, terpenes, polysaccharides (carbohydrates, uronic acids, sulphates), polyhydroxylated lactone, bioglycan, cyclic tripeptide, macrocyclic lactones *etc.* These marine constituents exert a desired pharmacological effect on body and thus act as natural immunomodulatory agents.

Conclusion

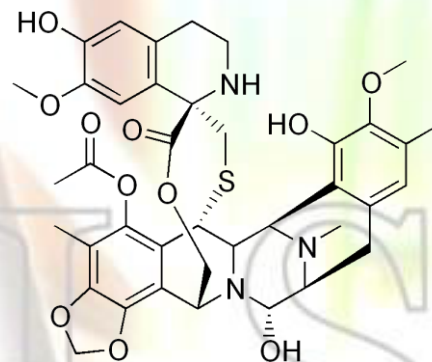
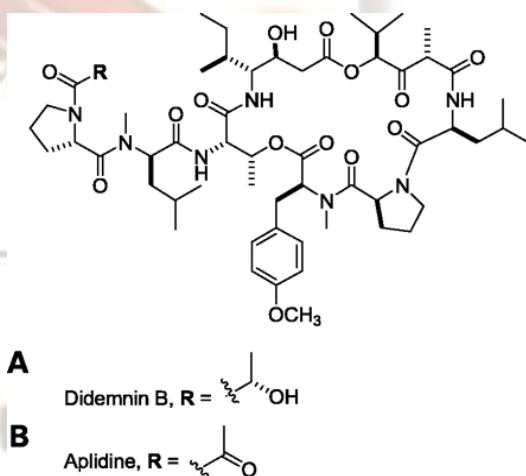
Marine have been a prime source of highly effective conventional drugs. Many studies have been performed to identify immunomodulator compounds with desired pharmacological activity and a limited toxicity. This review makes an attempt to give scientific account of use of valuable marine immunomodulatory sources. Some marine sources may stimulate the immune system like *Endarachne binghamiae*, *Gelliodes fibrosa*, *Sargassum ilicifolium*, *Phaeodactylum tricornutum* *etc* and some of them may suppress the immune responses example *Spirulina fusiformis*, *Chlorella stigmatophora*, *Trididemnum solidum*, *Euchelus asper* *etc.* Also the various secondary metabolites like Cyclic depsipeptide, terpenes, polysaccharides (carbohydrates, uronic acids, sulphates), polyhydroxylated lactone, bioglycan, cyclic tripeptide, macrocyclic lactones *etc.* have been exerting wide range of immunomodulating activity. Thus successful results have been achieved by following an appropriate screening approach.

References

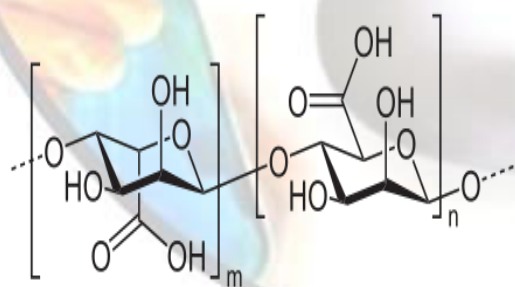
1. Kurokawa M., Shimizu T., Watanabe W. and Shiraki K. (2010). Development of New Antiviral Agents from Natural Products. *The Open Antimicrobial Agents Journal*, **2**:49-57.
2. Mancini I., Defant A. and Guella G. (2007). Recent Synthesis of Marine Natural Products with Antibacterial Activities. *Anti-Infective Agents in Medicinal Chemistry*, **6**:17-48.
3. Fahmy H., Khalifa S.I., Konoshima T. and Zjawiony J.K. (2004). An Improved Synthesis of 7, 8-Epoxy-1, 3, 11-cembratriene- 15 R(α), 16-diol, a Cembranoid of Marine Origin with a Potent Cancer Chemopreventive Activity. *Mar. Drugs*, **1**:1-7.
4. Jain R. (2008). Marine organisms: Potential source for drug discovery. (2008). *Current science*, **94**:292.
5. Davis G.D.J. and Vasanthi A.H.R. (2011). Seaweed metabolite database (SWMD): A database of natural compounds from marine Algae. *Bioinformation*, **5**:361-364.
6. Murti Y. and Agrawal T. (2010). Marine derived pharmaceuticals-Development of natural health products from marine biodiversity. *International Journal of ChemTech Research*, **2**: 2198-2217.
7. Agarwal S. and Singh V.K. (1999). Immunomodulators: A review of studies on Indian medicinal plants and synthetic peptides. Part 1: Medicinal plants. *PINSA*, **B 65**(3-4): 179-204.
8. Billiau A. and Matthys P. (2001). Modes of action of Freund's adjuvants in experimental models of autoimmune diseases. *Journal of Leukocyte Biology*, **70**: 850-860.
9. Ford M Susan. and Roach S Sally. (2009). Introductory Clinical Pharmacology 27th Ed. *Lippincott Williams and Wilkins, USA*, 567-568.
10. El-Sheikh ALK (2008). Renal transport and drug interactions of Immunosuppressants. *Radboud University Nijmegen*, 62.
11. Constantine S. Mitsiades., Enrique M. Ocio., Atanasio Pandiella., et al. (2008). Aplidin, a Marine Organism-Derived Compound with Potent Antimyeloma Activity In vitro and In vivo. *Cancer Research*, **68**: 5216-5225.
12. Bourguet-Kandracki M.L., Debitus C. and Guyot M. (1996). Dipuuphedione, a Cytotoxic New Red Dimer from a New Caledonian Marine Sponge *Hyrtios* sp. *Termhedron Letters*, **37**:3861-3864.

13. Rathod D.B., Lahiri S, Yadav G.K. and Shah M.B. (2010). Immunomodulatory and Antioxidant Activity of *Curculigo orchoides* Gaertn. *International Journal of PharmTech Research*, **2**:1197-1203.
14. Allavena P., Signorelli M., Chieppa M., Erba E., Bianchi G., Marchesi F., Olimpico CO., Bonardi C., Garbi A., Lissoni A., Braud F., Jimeno J. and D'Incalci M. (2005). Anti-inflammatory properties of the novel antitumor agent yondelis (trabectedin): inhibition of macrophage differentiation and cytokine production. *Cancer Res.*, **65**: 2964-71.
15. Huang R. and Lee H.T. (2005). Immunological Properties of the Marine Brown Alga *Endarachne binghamiae* (Phaeophyceae). *International Journal of Applied Science and Engineering*, **3**: 167-173.
16. Chandraraj S., Prakash B. Navanath K. (2010). Immunomodulatory activities of ethyl acetate extract of two marine sponges *Gelliodes fibrosa* and *Tedania anhelans* and brown algae *Sargassum ilicifolium* with reference to phagocytosis. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*, **2**:302-307.
17. Rifai S., Fassouane A., Pinho P.M., Kijjoa A., Nazareth N., Nascimento M.S.J. and Herz W. (2005). Cytotoxicity and Inhibition of Lymphocyte Proliferation of Fasciculatin, a Linear Furanosesterterpene Isolated from *Ircinia variabilis* Collected from the Atlantic Coast of Morocco. *Mar. Drugs*, **3**: 15-21.
18. Kim K.H., Kim Y.W., Kim H.B., Lee B.J. and Lee D.S. (2006). Anti-apoptotic activity of laminarin polysaccharides and their enzymatically hydrolyzed oligosaccharides from *Laminaria japonica*. *Biotechnol Lett.*, **28**: 439-46.
19. Fast M.D., Johnson S.C., Eddy T.D., Pinto D. and Ross N.W. (2007). *Lepeophtheirus salmonis* secretory/excretory products and their effects on Atlantic salmon immune gene regulation. *Parasite Immunology*, **29**: 179-189.
20. Campa-Cordova A.I., Hernandez-Saavedra N.Y. and Ascencio F. (2002). Superoxide dismutase as modulator of immune function in American white shrimp (*Litopenaeus vannamei*). *Comp Biochem Physiol C Toxicol Pharmacol.*, **133**: 557-65.
21. Akerkar A.S., Ponkshe C.A. and Indap M.M. (2009). Evaluation of immunomodulatory activity of extracts from marine animals. *Indian Journal of Marine Sciences*, **38**:22-27.
22. Karnjanapratum S. and You S. (2010). Molecular characteristics of sulfated polysaccharides from *Monostroma nitidum* and their in vitro anticancer and immunomodulatory activities. *Int J Biol Macromol.*
23. Selvin J., Huxley A.J. and Lipton A.P. (2004) Immunomodulatory potential of marine secondary metabolites against bacterial diseases of shrimp. *Aquaculture*, **230**: 241-248.
24. Roy M.C., Ohtani I.I., Ichiba T., Tanaka J., Satari R. and Higa T. (2000). New Cyclic Peptides from the Indonesian Sponge *Theonella swinhoei*. *Tetrahedron*, **56**: 9079-9092.
25. Okai Y., Higashi-Okai K., Ishizaka S., Ohtani K., Matsui-Yuasa I. and Yamashita U. (1998). Possible immunomodulating activities in an extract of edible brown alga, *Hijikia fusiforme* (Hijiki). *Journal of the Science of Food and Agriculture*, **76**: 56-62.
26. Joung Han Yim, Eunwha Son, Suhkneung Pyo and Hong Kum Lee. (2005). Novel sulfated polysaccharide derived from red-tide microalga *Gyrodinium impudicum* strain KG03 with immunostimulating activity in vivo. *Marine Biotechnology*, **7**: 331-338.
27. Itoh H, Noda H, Amano H, Zhuaug C, Mizuno T. and Ito H. (1993). Antitumor activity and immunological properties of marine algal polysaccharides, especially fucoidan, prepared from *Sargassum thunbergii* of Phaeophyceae. *Anticancer Res.*, **13**: 2045-52.
28. Shan B. E., Yoshida Y., Kuroda E. and Yamashita U. (1999). Brief Communication Immunomodulating activity of seaweed extract on human lymphocytes in vitro. *International Journal of Immunopharmacology*, **21**: 59-70.
29. Zhou G., Sun Y.P., Xin H., Zhang Y., Li Z. and Xu Z. (2003). In vivo antitumor and immunomodulation activities of different molecular weight lambda-carrageenans from *Chondrus ocellatus*. *Pharmacological Research*, **50**: 47-53.
30. Vaeck M., Grooten J., Hamers R. and De Baetselier P. (1983). The immunomodulatory effect of anti-*Micrococcus luteus* antibodies I. Effect on in vitro rabbit T cell functions. *European Journal of Immunology*, **13**: 772-778.
31. Kijjoa A. and Sawangwong P. (2004). Drugs and Cosmetics from the Sea. *Mar. Drugs*, **2**: 73-82.

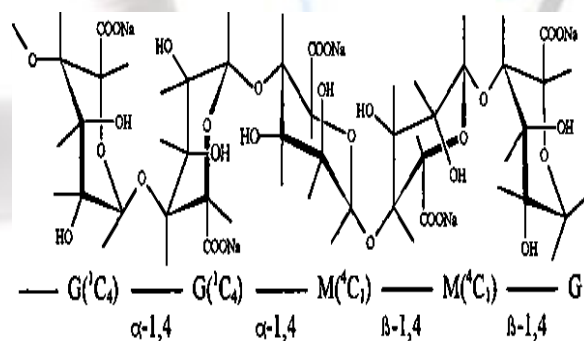
32. Villa F.A., Lieske K. and Gerwick L. (2010). Selective MyD88-dependent pathway inhibition by the cyanobacterial natural product malyngamide F acetate. *European Journal of Pharmacology*, **629**: 140-146.
33. Ciasullo L., Casapullo A., Cutignano A., Bifulco G., Debitus C., Hooper J., Gomez-Paloma L. and Riccio R. (2002). Renieramide, a Cyclic Tripeptide from the Vanuatu Sponge *Reniera* n. sp. *J. Nat. Prod.*, **65**: 407-410.
34. Ovodova R.G., Glazkova V.E., Mikheyskaya L.V., Molchanova V.I., Isakov V.V., Ovodov Y.S. and Molina L.E.F. (1992). The structure of mytilan, a bioglycan-immunomodulator isolated from the mussel *Crenomytilus grayanu*. *Carbohydrate Research*, 221-226.
35. Guzman S., Gato A., Lamela M., Freire-Garabal M. and Calleja, J. M. (2003), Anti-inflammatory and immunomodulatory activities of polysaccharide from *Chlorella stigmatophora* and *Phaeodactylum tricornutum*. *Phytotherapy Research* **17**: 665-670.
36. Fangmei Yang, Ying Shi, Jianchun Sheng and Qiuhui Hu. (2005). *In vivo* immunomodulatory activity of polysaccharides derived from *Chlorella pyrenoidosa*. *European Food Research and Technology*, **224**: 225-228.
37. Rasool M.K. and Sabina E.P. (2008). Appraisal of immunomodulatory potential of *Spirulina fusiformis*: an *in vivo* and *in vitro* study. *Journal of Natural Medicines*, **63**: 169-175.



Trabectedin

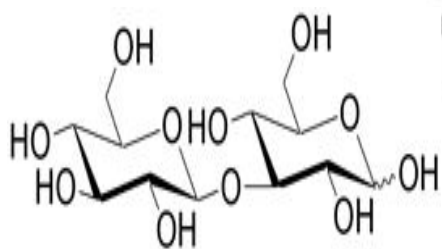


Alginate acid

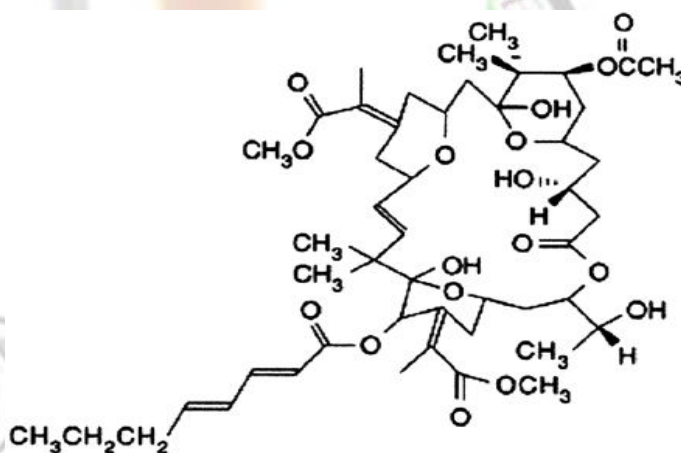
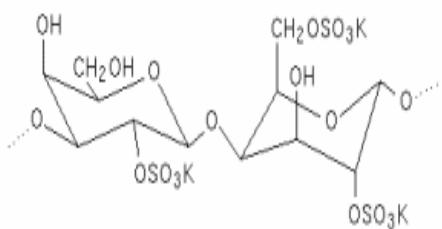
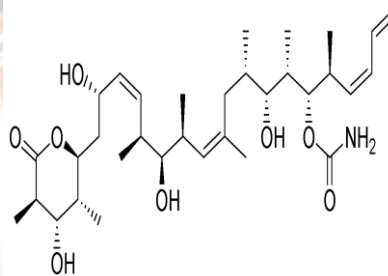


Sodium alginate

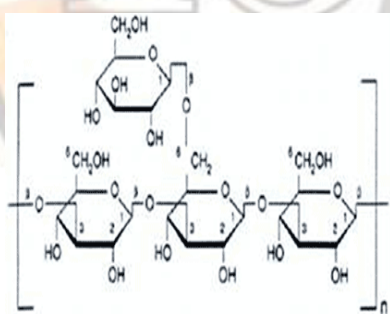
Laminarin



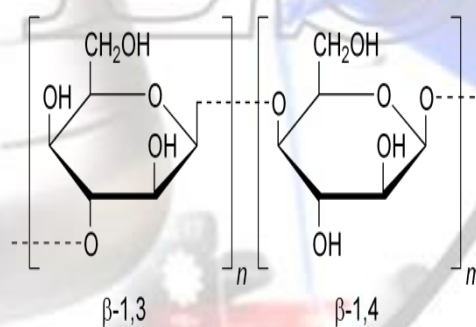
Discodermolide



Carrageenan



Bryostatins



Polysaccharide

 β -Glucan

Fig. 1: Chemical structures of reported marine immunomodulators

Table 1: Showing various marine immunomodulators¹¹⁻³⁷

Marine source	Chemical constituents	Immunomodulatory action
<i>Aplidium albicans</i> , Polyclinidae	Cyclic depsipeptide (Aplidin)	Aplidin caused <i>in vivo</i> anti-MM (multiple myeloma) activity in a xenograft plasmacytoma murine model
<i>Hyrrios erecta</i> , Thorectidae	Puupehedione, dipuupehedione, bispuupehenone	Puupehedione exerted a desired immunomodulatory potential
<i>Curculigo orchoides</i> , Hypoxidaceae	3,5-diacetoxy 2-methoxy 6-methyl phenyl acetic acid	Methanol extract seemed to work on both specific and non-specific immune pathways, whereas polysaccharide rich aqueous extract appeared to stimulate proinflammatory immune responses
<i>Ecteinascidia turbinata</i> , Perophoridae	Yondelis (Trabectedin)	Yondelis caused selective decrease of monocyte counts and of <i>ex vivo</i> macrophage differentiation
<i>Endarachne binghamiae</i> , Phaeophyceae	Polysaccharides (sodium alginate, alginic acid), glycoprotein	Sodium alginate exhibited strong stimulation activity for macrophage and T cell proliferation and also alginic acid but to a lesser extent. A glycoprotein isolated from the reported alga was also a strong proliferation stimulant. Additionally, it significantly induced the production of TNF- α and nitric oxide by macrophages and IFN- γ by T cells in a concentration-dependent manner. These assay results suggested that alginate and protein of the reported alga could be promising immune stimulants and modulators
<i>Gelliodes fibrosa</i> , Niphatidae	Terpenes, steroids and lipids	The ethyl acetate extracts of <i>G. fibrosa</i> on <i>in vivo</i> carbon clearance test have shown moderate immunostimulatory effect
<i>Tedania anhelans</i> , Tedaniidae	Terpenes, steroids and lipids	The ethyl acetate extracts of <i>T. anhelans</i> on <i>in vivo</i> carbon clearance test have shown moderate immunostimulatory effect <i>In vitro</i> study revealed that <i>T. anhelans</i> stimulated chemotatic, phagocytic and intracellular killing of human neutrophils
<i>Sargassum ilicifolium</i> , Sargassaceae	Terpenes, steroids and lipids	<i>S. ilicifolium</i> have shown prominent immunostimulatory effect. <i>In vitro</i> study revealed that <i>S. ilicifolium</i> stimulated chemotatic, phagocytic and intracellular killing of human neutrophils
<i>Ircinia variabilis</i> , Irciniidae	Fasciculatin(sesterterpenes)	Fasciculatin showed a moderate cytotoxicity and no selectivity on the cancer cell lines. Immunomodulatory activity of Fasciculatin found that it was inactive on human lymphocyte proliferation
<i>Laminaria japonica</i> , Laminariaceae	Laminarin oligosaccharides, polysaccharides	A mouse cDNA microarray showing the genes coding for immune response proteins were induced and apoptotic cell death proteins were reduced significantly by LO (Laminarin oligosaccharides) provided preliminary information regarding the immunomodulatory mechanism of LO

<i>Lepeophtheirus salmonis</i> , Caligidae	Trypsins	<i>L. salmonis</i> caused inhibitory effect on one centrally involved inflammatory gene (IL-1 β)
<i>Litopenaeus vannamei</i> , Penaeoidea	Polysaccharides	Results indicated that the immunomodulatory action of superoxide dismutase (SOD) and its possible use as an indicator of immune responses
<i>Nematopaleamon tenuipes</i> , Penaeidae	Fractions of Petroleum ether: ethyl acetate (1:1) containing constituents	The Delayed type Hypersensitive reaction assay showed stimulation, whereas in the PFC(Plaque Forming Cell) assay it showed immunosuppression
<i>Hemifusus pugilinus</i> , Melongenidae	Fractions of Petroleum ether: ethyl acetate (1:1) containing constituents	The Delayed type Hypersensitive reaction assay showed stimulation. In the PFC assay it showed immunostimulation
<i>Rastrelliger kanagurta</i> , Scombridae	Fractions of Petroleum ether: ethyl acetate (1:1) containing constituents	Fractions showed stimulation in Delayed type Hypersensitive reaction assay. In the PFC assay it showed immunostimulation
<i>Euchelus asper</i> , Trochidae	Fractions of Petroleum ether: ethyl acetate (1:1) containing constituents	Fractions were less effective in the Delayed type Hypersensitive reaction assay showed stimulation. In the PFC assay it showed immunosuppression
<i>Monostroma nitidum</i> , Gomontiaceae	Sulfated polysaccharides (carbohydrates, uronic acids, sulphates)	Polysaccharides stimulated a macrophage cell line, Raw 264.7 cells, inducing considerable NO (nitric oxide) and PGE(2), production, which suggested that they could be strong immunomodulators
<i>Dendrilla nigra</i> , Dysideidae	Lipopolysaccharides	<i>Dendrilla</i> medicated group exhibited enhanced phagocytosis against <i>E. coli</i>
<i>Ulva fasciata</i> , Ulvaceae	Lipopolysaccharides	<i>Ulva</i> diet significantly increased the defense factors such as haemogram, agglutination index, phagocytic rate, bacterial clearance and serum bactericidal activity of treated shrimps over the control group
<i>Theonella swinhoei</i> , Theonellidae	Cyclic peptides, barangamides B, C, and D and a new depsipeptide, theonellapectolide	Theonellapectolides showed mild immunosuppressive activity
<i>Hijikia fusiforme</i> , Sargassaceae	Polysaccharide	Enhancing activity for the proliferative response of spleen cells in endotoxin-nonresponder C3H/HeJ mice
<i>Gyrodinium impudicum</i> , Picornaviridae	Polysaccharide	<i>G. impudicum</i> showed immunostimulatory effects and enhanced the tumoricidal activities of macrophages and NK cells <i>in vivo</i>
<i>Sargassum thunbergii</i> , Sargassaceae	Polysaccharide (Fucoidan)	Fucoidan enhanced the phagocytosis and chemiluminescence of macrophages
<i>Meristotheca papulosa</i> , Solieriaceae	Polysaccharide	Extracts of <i>M. papulosa</i> (green) markedly stimulated human lymphocytes to proliferate
<i>Chondrus ocellatus</i> , Gigartinaceae	Carrageenans	In this study, tumor-inhibiting activities, weight of immune organ, nature killer cells activity, lymphocyte proliferation ratio and pathological slice of spleen and tumor cells from the control group and λ -carrageenan-treated mice of transplanted S180 and H22 tumor were investigated. The results indicated that the five λ -carrageenan samples showed antitumor and immunomodulation activities

<i>Micrococcus luteus</i> Micrococcaceae	Anti- <i>Micrococcus luteus</i> antibodies	Selective T cell immunosuppressive potential
<i>Bugula neritina</i> , Bugulidae	Macrocyclic lactones (Bryostatins)	Bryostatin 1 was found to bind to protein kinase C with high affinity, which may be the mechanistic basis for both observed anticancer and immunostimulating activities
<i>Trididemnum solidum</i> , Didemnidae	Depsipeptides (Didemnin B)	<i>T. solidum</i> extract showed immunosuppressive activities
<i>Discodermia</i> species, Theonellidae	Polyhydroxylated lactone (Discodermolide)	<i>Discodermia</i> showed immunosuppressive and cytotoxic activity
<i>Lyngbya majuscula</i> , Oscillatoriaceae	Peptides (Microcolin-A)	<i>L. majuscula</i> showed inhibitory effect against innate immune system (immunosuppressive)
<i>Reniera</i> species, Chalinidae	Cyclic Tripeptide (Renieramide)	<i>Reniera</i> showed immunomodulating activity in preliminary tests
<i>Crenomytilus grayanus</i> , Mytilidae	Bioglycan (Mytilan)	Mytilan isolated from the mantle of the mussel <i>C. grayanus</i> possessed high immunomodulating activity
<i>Chlorella stigmatophora</i> Syngnathidae	Polysaccharide	<i>C. stigmatophora</i> extract showed immunosuppressant effects
<i>Phaeodactylum tricorutum</i> , Phaeodactylaceae	Polysaccharide	<i>P. tricorutum</i> extract showed immunostimulatory effects
<i>Chlorella pyrenoidosa</i> , Mytilidae	Polysaccharide	Oral administration of <i>C. pyrenoidosa</i> polysaccharides, significantly enhanced phagocytic rate and phagocytic index
<i>Spirulina fusiformis</i> , Oscillatoriaceae	Polysaccharide	<i>S. fusiformis</i> showed immunosuppressive effect