



A Review on Immunomodulatory Medicinal Plants

Aarti Dubey^{1*} and Vishal Gupta²

1, Research Scholar, Faculty of Pharmacy

2, Dean Faculty of Pharmacy, Mansarovar Global University, Sehore, (M.P.) - India

Article info

Received: 04/10/2021

Revised: 20/11/2021

Accepted: 29/11/2021

© IJPLS

www.ijplsjournal.com

Abstract

The ayurvedic system of medicine is one of the oldest systems of medicine and includes various ethnopharmacological activities such as immunostimulation, tonic, neurostimulation, anti-ageing, antibacterial, antiviral, antirheumatic, anticancer, adaptogenic, etc. An entire section of the Materia Medica of Ayurveda is devoted to "Rasayana", drugs reputed to enhance body resistance. Listed as a class in the texts of traditional Indian Medicine literature, Rasayana consists of a number of plants reputed to promote physical and mental health, improve defense mechanisms of the body and enhance longevity. These attributes are similar to the modern concept of adaptogenic agents, which are known to afford protection of the human physiological system against diverse stressors. A number of medicinal plants as Rasayanas have been claimed to possess immunomodulatory activity. The present review highlights the use of various medicinal plants as immunomodulators.

Key-words: Ayurveda, Medicinal plants, Immunomodulators.

Introduction

Medicinal plants have tremendous potential to yield novel compounds as model for future drugs. One of the fast developing area of drug development involves the search for novel immunomodulatory agents having either immune-stimulatory or immune-suppressant activity that could be used for the treatment of various immune dysfunctions or in case of residual cancer. The intralesional application of BCG and systemic use of chemical non-specific immunomodulator levamisole have shown promising results, however, break-through in the field of immunomodulators was achieved when a strong immunosuppressant drug, cyclosporine was discovered that prevented the rejection of graft and had use in other auto-immune diseases. Immu-21 a polyherbal immunomodulator has been reported to promote host immune functions by augmenting both arms of the immune system

i.e. humoral and cell-mediated immunity and has been reported to be safe and non-toxic. There are a number of medicinal plants whose crude preparations, formulations or purified compounds have shown strong immunomodulatory activity. [1-4]

*Corresponding Author

Medicinal Plants as Immunomodulators [7-31]

Curcuma longa

Curcumin, an active ingredient of *Curcuma longa* ('Haldi' or Turmeric, Family: Zingiberaceae) and powdered root extract from *W. somnifera* (20 mg/dose/animal, i.p) were found to increase total WBC count, increase in circulating antibody titre against Sheep Red Blood Corpuscles (SRBC), splenic plaque forming cells (PFC), bone marrow cellularity, alpha esterase positive cells and an increase in the phagocytic activity of macrophages in BALB/c mice. The petroleum ether extract of the rhizomes of *C. longa* also showed significant anti-inflammatory activity and was effective in delayed hypersensitivity.

Piper betle Linn.

It is a widely distributed plant in the tropical and subtropical regions of the world and has been attributed as traditional herbal remedy for many diseases. *P. betle* has several landraces. The *in vivo* immunomodulatory efficacy in the crude methanolic extract and its n-hexane, chloroform, n-butanol fractions of the female plant at various doses in murine model. The crude methanol extract and n-hexane fraction were found to potentiate both the humoral (PFC and hemagglutination titre) and cellular (lymphoproliferation, macrophage activation, delayed type hypersensitivity) immune responses in mice. The splenocytes of treated mice displayed enhanced population of CD4+ and CD8+ T cells as well as CD19+ B cells. The n-hexane fraction induced a biased type 2 cytokine response (increased IL-4, decreased IFN γ) while the chloroform fraction produced a predominant type 1 cytokine response. On the other hand, the crude methanolic extract possessed a mixed T_H cell response suggesting a remarkable immunomodulatory property in this plant.

Nepeta ucrainica L.

Phytochemical investigations of the aerial parts of the plant *Nepeta ucrainica* L. (used as herbal tea in Kazakhstan) resulted in the isolation of verbascoside. The *in vitro* immunomodulatory activity of verbascoside was investigated by assessing neutrophil function, chemotaxis and intracellular killing activity. Verbascoside showed an increased chemotactic activity and had a positive effect on respiratory burst of neutrophils.

The opposite effect was observed with increasing doses demonstrating a possible suppression of neutrophil intracellular killing activity.

Tinospora cordifolia

Also known as Guduchi belongs to family Menispermaceae has been reported to contain a polysaccharide as an immunomodulatory constituent in its crude stem extract and was found to impart its effect as an adjuvant to augment process of ulcer healing, bacterial eradication and polymorphonuclear phagocytosis.

Alpinia galanga

Another polysaccharide isolated from the rhizome of *Alpinia galanga* showed a marked stimulant effect on the reticulo-endothelial system (RES) and increased the cellularity in the peritoneal cavity as well as in spleen after treatment of mice. "Ginseng", a well known herbal remedy, developed mainly from the roots of Korean or *Panax ginseng*

Asian ginseng (*Panax ginseng*), Siberian ginseng (*Eleutherococcus senticosus*), or American ginseng (*Panax quinquefolius*), belong to family Araliaceae was reported to impart its immunomodulatory functions by increasing antibody concentration and natural killer (NK) cell activity in the treated individuals. It has also been reported to confer a lower incidence of influenza and colds. The main active agent in *Panax ginseng* is identified as ginsenoside, a triterpene saponin.

Mollugo verticillata L.

An independent study with ethanolic extracts of *Mollugo verticillata* L. (Family: Molluginaceae) has confirmed stimulation of peritoneal macrophages and increased production of nitric oxide (NO), however, if used along with BCG or *Mycobacterium tuberculosis* antigen, it suppressed the activity of these cells. An aqueous extract prepared from the fruits of the plant *Emblica officinalis*

Also *Phyllanthus emblica*; Family: Euphorbiaceae has been shown to possess antioxidant properties and protected mice from the chromosome-damaging effects of the well known carcinogen 3, 4-benzo (a) pyrene thereby increasing the life span of tumor bearing mice. Immunomodulatory effects of the aqueous extract of the tea, *Camellia sinensis*, was also reported *in vitro* in the form of enhanced production of neopterin, a sensitive

marker of cell-mediated immunity (CMI) in unstimulated human peripheral mononuclear cells, however, a reduction of neopterin formation was observed when cells were stimulated with mitogens.

Centella asiatica

Another important traditional medicinal plant, *Centella asiatica* (Family: Apiaceae) commonly known as "Mandukaparani" in India, is used in folk medicine as rasayana. The alcoholic extract of *C. asiatica* has been reported to have reticuloendothelial stimulating activity and increases the antibody titer and delayed type hypersensitivity (DTH) response when fed at 100 mg/ kg body weight. The active constituent of *C. asiatica* was identified as Asiaticosides.

Azadirachta indica

The Indian Vedic plant Neem (*Azadirachta indica*, Family: Meliaceae) is also a powerful immunostimulatory plant reported to be non-toxic and hematostimulatory.

Andographis paniculata

The methanolic extract of *Andographis paniculata* (Family: Acanthaceae), also known 'Kaal Megh, has been reported to contain andrographolide, 14-deoxyandrographolide and 14-deoxy-11, 12-didehydroandrographolide as active constituents which are attributed with the strong immunostimulatory properties. These compounds enhanced the cellular proliferation and interleukin-2 (IL-2) production in human peripheral blood lymphocytes and reversed the cyclophosphamide induced immunosuppression.

Chlorophytum borivillianum

A polysaccharide fraction derived from hot water extraction of *Chlorophytum borivillianum* (Family: Liliaceae) comprising ~31% inulin-type fructans and ~25% acetylated mannans was found to stimulate NK cell activity *in vitro*, however, *in vivo* evaluation of the aqueous extract was found to strengthen the humoral immune functions of Wistar strain of albino rats.

Prominent work of herbal Medicine [32-50]

A study conducted by Dhuley with plants *Asparagus racemosus*, *Tinospora cordifolia*, *Withania somnifera* and *Picrorhiza kurroa* revealed significant attenuation of ochratoxin carcinogen induced suppression of chemotactic activity as well as IL-1 and TNF α production by macrophages of treated mice. These plants also

produced leucocytosis with predominant neutrophilia and prevented, to varying degrees, the leucopenia induced by cyclophosphamide. The plant *A. racemosus* induced excessive production of TNF α while *W. somnifera* potentiated macrophage chemotaxis, NO production over that of controls. Immunostimulatory effects have also been reported from the bark aqueous extract of *pule* (*Alstonia scholaris*, Family: Apocynaceae) in BALB/c mice. The extract stimulated non specific immune response, restored the prednisolone induced reduction of phagocytic activity and protected the body from opportunistic infection caused by *Escherichia coli*. The hexane and aqueous extracts/ fractions of *Carica papaya* seeds significantly enhanced the proliferation of lymphocytes towards phytohemagglutinin (PHA) sensitization and inhibited the classical complement-mediated hemolytic pathway. Among the numerous plants known for their traditional medicinal use, *Pterospermum acerifolium* commonly known as Kanak Champa was also reported to have hepatoprotective activity. Preliminary phytochemical analysis of the leaf extract of this plant revealed the presence of several compounds like steroids, flavonoids, alkaloids and glycosides. We have reported a contrasting immunomodulatory activity in different vegetative parts of this plant i.e. a strong immunosuppressive activity in the seed part and an excellent immunostimulatory activity in the floral ethanolic extract derived from this plant demonstrating the diverse pharmacological activity in different parts of the same plant. *Withania coagulans* has been ascribed to possess various biological activities including anti-hyperglycemic activity. This plant is known to contain withanolides (withaferin-A), phenolic tannin, flavonoids and flavonols. Shah and Juvekar have reported the macrophage activating effects in the form of up-regulated production of nitric oxide and augmentation in the antibody levels of animals treated with an aqueous extract from the leaves of *Murraya koenigii*, commonly known as curry leaf tree (Family: Rutaceae). *M. koenigii* extract has also been reported to have anti-hyperglycemic and cholesterol lowering effects. Bafna and Mishra described the *in vitro* antioxidant and immunomodulatory activity in mice treated with the alkaloidal fraction from the

roots of *Cissampelos pareira* Linn in a dose dependent manner. The alkaloid fraction was found to have significant immunosuppressive activity at lower doses (25 and 50 mg/kg) while no activity was observed at higher doses (75 and 100 mg/kg) revealing the immunosuppressive and antioxidant activities in the alkaloidal fraction of *C. pareira* roots. Khanfor the first time have reported immunomodulatory activity in the ethanol (50%) extracts of the seed and root of *Nyctanthes arbor-tristis* L. against systemic candidiasis in mice. Sharififar demonstrated immunomodulatory activity of the aqueous extract from the fruits of *Heracleum persicum* Desf. (Apiaceae) at 50, 100 and 200 mg/kg in female Swiss mice. The extract was found to stimulate both humoral and cellular arms of the immune system. The alkaloids, tetrrandrine found in the roots of *Cyclea arnotii* Miers (Family: Menispermaceae) and rohitukine present in the stem bark of *Dysoxylum binectariferum* Hook. f. (Family: Meliaceae) also possessed immunomodulatory properties and plant has been indicated in the treatment of various immune disorders.

The hexane, chloroform and ethanol extracts of *Boerhavia diffusa* (Commonly known as Punarnava, Family: Nyctaginaceae) were evaluated *in vitro* for their immunomodulatory activities on human peripheral blood mononuclear cells (PBMCs). The hexane extract was found to stimulate proliferation of PBMCs, LPS induced NO production and, PHA/ LPS induced IL-2 and TNF α production. The pure compound Bd-I (eupalitin-3-O-h-D galactopyranoside) purified from the ethanolic extract inhibited production of PHA stimulated IL-2 at the protein and mRNA transcript level and LPS stimulated TNF α production in human PBMCs at equivalent dose. This compound also blocked the activation of DNA binding of nuclear factor- κ B and AP-1, the two major transcription factors centrally involved in expression of IL-2 and IL-2R gene, which are necessary for the activation and proliferation of T cells.

Conclusion

Plants have served a great deal in treating various diseases and maintaining good health. There is huge information available on several plants and each plant is having several traditional and

scientifically established therapeutic uses. Several researchers have reviewed literature on several categories of drugs and medicinal plants. This list of this literature survey is restricted to the conventional and historical uses and its therapeutic effect on the physiology has been reported in reputed journals on the plants used in this study. Several review articles, research papers, text books, and databases are referred over a wide period. The literature available on the marketed immunomodulators used in this study.i.e. various Ayurvedic Marketed to increase immunity.

References

1. Archana. , Suresh Jatawa., Rajkumar Paul and Archana Tiwari.,(2011). Indian medicinal plants:A rich source of natural immunomodulator,vol-7,issue-2 page no-198-205.
2. Agarwal, S.S. and V.K. Singh., (1999). Immunomodulators: A review of studies on Indian medicinal plants and synthetic peptides. Part 1: Medicinal plants. Proc. Indian Natl. Sci. Acad. Part B: Biol. Sci., B65: 179-204.
3. Bafna A. R., and Mishra A. R., Protective effect of bioactive fraction of *Sphaeranthus indicus* Linn. against cyclophosphamide induced suppression of humoral immunity in rat, Journal of Ethnopharmacology, 2006, 104, 1-4. (ref-45)
4. Bhutani, K.K. and V.M. Gohil., (2010). Natural products drug discovery research in India: Status and appraisal. Indian J. Exp. Biol., 48: 199-207.
5. Ansari, J.A. and N.N. Inamdar., (2010). The promise of traditional medicines. Int. J. Pharmacol., 6: 808-812.
6. Chih-Chun Wen, Hui-Ming Chen, and Ning-Sun Yang(2012) Developing Phytocompounds from Medicinal Plants as Immunomodulators Adv Bot Res 2012;62:197-272. doi: 10.1016/B978-0-12-394591-4.00004-0. Epub 2012 Jun 19.
7. Claeson, U.P., T. Malmfors., G. Wikman and J.G. Bruhno., (2000). *Adhatoda vasica*: A critical review of ethnopharmacological and toxicological data. J. Ethnopharmacol., 72: 1-20.
8. Dahanukar, S.A., R.A. Kulkarni and N.N. Rege.,(2000). Pharmacology of medicinal plants and natural products. Indian J. Pharmacol., 32: 81-118.

9. De Pasquale, A.,(1984). Pharmacognosy: The oldest modern science. *J. Ethnopharmacol.*, 11: 1-16.
10. Desai, V.R., R. Ramkrishnan., G.J. Chintalwar and K.B. Sainis.,(2007). Gl-4A, an immunomodulatory polysaccharide from *Tinospora cordifolia*, modulates macrophage responses and protects mice against lipopolysaccharide induced endotoxic shock. *Int. Immunopharmacol.*, 7: 1375-1386.
11. Devasagayam, T.P.A., J.P. Kamat and N. Sreejayan.,(2001). Antioxidant Action of Curcumin. In: *Micronutrients and Health: Molecular Biological Mechanisms*. Nesaretnam, K. and L. Packer (Eds.). AOCS Press, Champaign, IL, USA., pp: 42-59.
12. Dixit, P., S. Ghaskadbi., H. Mohan and T.P. Devasagayam.,(2005). Antioxidant properties of germinated fenugreek seeds. *Phytother. Res.*, 19: 977-983.
13. Dinesh Kumar., Vikrant Arya., Ranjeet Kaur., Zulfiqar Ali Bhat., Vivekkumar Gupta (2012). A review of immunomodulators in the Indian traditional health care system. *Journal of Microbiology, Immunology and Infection* Volume 45, Issue 3, Pages 165-184.
14. F Sharififar., S Pourmourmohammadi., M Arabnejad.,(2009) . nopr.niscair.res.in
15. G.R. Arun Raj., UShailaja., Prasanna N. Raos., and S. Ajayan.,(2014). Department of PG Studies in Kaumara Bhritya, Shri Dharmasthala Manjunatheshwara College of Ayurveda and Hospital, Hassan, Tamil Nadu, India
16. Ganiu, L., D. Karan., S. Chanda., K.K. Srivastava., R.C. Sawhney and W. Selvamurthy., (2003). Immunomodulatory effects of agents of plant origin. *Biomed. Pharmacother.*, 57: 296-300.
17. Ghule B.V.,(2003). Preliminary phytochemical screening and pharmacological investigation of Myristica fragrance for immunomodulatory activity; A thesis submitted in Nagpur University, Nagpur, (ref- 26)
18. Goldfrank, L., N. Lewin, N. Flomenbaum and M.A. Howland, 1982. The pemicious panacea: Herbal medicine. *Hosp. Physician*, 18: 64-69.
19. Guerra, R.N., H.A. Pereira., L.M. Silveria and R.S. Olea.,(2003). Immunomodulatory property of *Alternanthera tenella* Colla aqueous extracts in mice. *Braz. J. Med. Biol. Res.*, 36: 1215-1219.
20. Hemant sagrawat and Md. Yaseen Khan (2007) Immunomodulatory Plants: A Phytopharmacological Review. Research Associate, Getz Pharma Research Pvt. Ltd., Mumbai 2Department 248-260.
21. Hoareau, L. and E.J. DaSilva, 1999. Medicinal plants: A re-emerging health aid. *Electron. J. Biotechnol.*, 2: 56-70.
22. Ismail, S. and M. Asad.,(2009). Immunomodulatory activity of *Acacia catechu*. *Indian J. Physiol. Pharmacol.*, 53: 25-33.
23. J Vaghasiya., M Datani., K Nandkumar.,(2010) . Int J Pharm Biol-researchgate.net
24. Jachak, S.M. and A. Saklani., (2007). Challenges and opportunities in drug discovery from plants. *Curr. Sci.*, 92: 1251-1257.
25. K. L. Satpute1, M. M. Jadhav1, R. S. Karodil, Y. S. Katare, M. J. Patil , Rukhsana Rub and A. R. Bafna (2009) Immunomodulatory activity of fruits of *Randia dumetorum* Lamk journal of Pharmacognosy and phytotherapy.
26. Kalia, A.N., (2005). Text Book of Industrial Pharmacognosy. Oscar Publication, New Delhi, India.
27. Kulkarni, P., R. Paul and N. Ganesh., (2010). In vitro evaluation of genotoxicity of avocado (*Persea Americana*) fruit and leaf extracts in human peripheral lymphocytes. *J. Environ. Sci. Health C Environ. Carcinog. Ecotoxicol. Rev.*, 28: 172-187.
28. Kulkarni, S.D., J.C. Tila.k, R. Acharya., N.S. Rajurkar., T.P. Devasagayam and A.V. Reddy., (2006). Evaluation of the antioxidant activity of wheatgrass (*Triticum aestivum* L.) as a function of growth under different conditions. *Phytother. Res.*, 20: 218-227.
29. Kumar, B., M. Vijayakumar., R. Govindarajan and P. Pushpangadan.,(2007). Ethnopharmacological approaches to wound healing: Exploring medicinal plants of India. *J. Ethnopharmacol.*, 114: 103-113.
30. Mehrotra, S., K.P. Mishra., R. Maurya., R.C. Srimal and V.K. Singh.,(2002). Immunomodulation by ethanolic extract of *Boerhaavia diffusa* roots. *Int. Immunopharmacol.*, 2: 987-996.
31. M.Suresh Gupta., H.N. Shivaprasad, M.D. Kharya&A.C. Rana immunomodulatory Activity of the Ayurvedic Formulation online publication.
32. Mehrotra, S., K.P. Mishra., R. Maurya., R.C. Srimal., V.S. Yadav., R. Pandey and V.K.

Singh., (2003). Anticellular and immunosuppressive properties of ethanolic extract of *Acorus calamus* rhizome. *Int. Immunopharmacol.*, 3: 53-61.

33. Mungantiwar, A.A., A.M. Nair., U.A. Shinde., V.J. Dikshit., M.N. Saraf., V.S. Thakur and K.B. Sainis., (1999). Studies on the immunomodulatory effects of *Boerhaavia diffusa* alkaloidal fraction. *J. Ethnopharmacol.*, 65: 125-131.

34. Muthaliar, M., (1998). *Siddha Materia Medica* (Vegetable Section). Vol. I. 4th Edn., Tamilnadu Siddha Medical Council, Chennai.

35. Nagarathna P.K.M , Reena K, Sriram Reddy, Johnson Wesley(2013) Review on Immunomodulation and Immunomodulatory Activity of Some Herbal Plants *Int. J. Pharm. Sci. Rev. Res.*, 22(1), n° 41, 223-230

36. Nadkami. K.M.,(2000). *Indian Materia Medica*. Reprinted Vol. 1. Bombay Popular Prakashan, India, pp: 300-302..

37. Namrata Singh., MukulTailang and S.C. Mehta.,(2016). Department of Pharmacology G R Medical College, Gwalior, M. P. India.SOS in Pharmaceutitcal Sciences, Jiwaji University, Gwalior

38. Patwardhan, B., A.D.B. Vaidya and M. Chorghade.,(2004). Ayurveda and natural products drug discovery. *Curr. Sci.*, 86: 789-799.

39. Paul, R., P. Kulkarni and N. Ganesh.,(2010). Avocado fruit (*Persea americana* Mill.) exhibits chemo-protective potentiality against cyclophosphamide induced genotoxicity in *In-vitro* human lymphocyte culture. *Phytother. Res.*

40. Plaeger, F.S.,(2003). Clinical immunology and traditional herbal medicines. *Clin. Diagn. Lab. Immunol.*, 10: 337-338.

41. PranotiBelapurkar., Pragya Goyal And Preeti Tiwari-Barua., (2014).Department of Biotechnology, IPS Academy

42. Preeti Sharma., Pradeep Kumar, Rachna Sharma., Gaurav Gupta., Anuj Chaudhary .,(2017). *National Journal of Physiology, Pharmacy and Pharmacology*

43. Pushpangadan, P., (2006). *Important Indian Medicinal Plants of Global Interest*. Rajiv Gandhi Centre for Biotechnology, Kerala, India, pp: 95-109.

44. Raikumar, P., P. Murari and K.S. Nand.,(2010). Anticancer biology of neem (*Azadirachta Indica* L.): A mini review. *Anticancer Drugs*.

45. Ranjith, M.S., A.J.A. Ranjitsingh., S.G. Shankar., G.S. Vijayalaksmi., K. Deepa and H.S. Sidhu., (2008). Enhanced phagocytosis and antibody production by *Timospora cordifolia*: A new dimension in immunomodulation. *Afr. J. Biotechnol.*, 7: 81-85.

46. Rates, S.M.K., (2001). Plants as source of drugs. *Toxicon*, 39: 603-613.

47. Rinku. M.. V.V. Prasanth and G. Parthasarathy., (2009). Immunomodulatory activity of the methanolic extract of *Urena lobata* Linn. *Internet J. Pharmacol.*, Vol. 7.

48. S.A Bagwan., N.S Naikwade and J.Y Manure.,(2017) Department of Pharmacology, AppasahebBirnale College of Pharmacy, Sangli-416416, India.

49. Sagrawat H. and Khan Y.,(2007) Immunomodulatory Plants: A Phytopharmacological Review, *Pharmacognosy Reviews*, 1, 248-260.(ref-27)

50. Satakopan, S.,(1994). *Pharmacopeial standards for Ayurvedic, Siddha and Unani Drugs*. Proceedings of WHO Seminar on Medicinal Plants and Quality Control of Drugs Used in ISM. Ghaziabad, pp: 43.

Cite this article as:

Dube A. and Gupta V. (2021). A Review on Immunomodulatory Medicinal Plants. *Int. J. of Pharm. & Life Sci.*, 12(11):1-6.

Source of Support: Nil

Conflict of Interest: Not declared

For reprints contact: ijplsjournal@gmail.com